

Totten, R. E. and H. B. Howe, Jr. Enzyme profiler during synchronous development of conidiophore and conidia in *N. crassa*.

and Fincham 1968 *J. Bacteriol.* 95:1063), NAD- and NADP-glutamate dehydrogenase (Sanwal and Lata 1961 *Can. J. Microbiol.* 7: 319; Stine 1968 *J. Cell Biol.* 37:81), and NADH-oxidase (Stine 1968). We have used our method for obtaining synchronously-developing mycelial pads (1971 *Biochem. Genet.* 5:521) in examining the profiles of 12 enzymes to determine changes in their specific activities during development of conidiophore and conidia.

Such pads were incubated in petri dishes containing 10 ml of 1% Difco Bacto-Agar at 35°C under fluorescent illumination for 0, 1, 4, or 7 hr, washed with deionized water in a Buchner funnel, pressed dry, and then frozen. Each frozen pad was ground intermittently with a Virtis 45 homogenizer for 15 min with 5 g of glass beads in 25 ml of 0.05 M phosphate buffer (pH 7), and the homogenate was centrifuged at 15,000 x g for 30 min. The precipitate was resuspended in 5 ml of phosphate buffer and sonicated intermittently for 2 min, the sonicate was centrifuged at 15,000 x g for 30 min. and this supernatant was combined with the first supernatant. All procedures were carried out in the cold. Seven-hour pads had developed conidiophores and conidia; 4-hr pads, conidiophores, but not conidia; and 1-hr pads, neither of these structures. Dry weights (mg) and total soluble protein (mg) of the pads at 0, 1, 4, and 7 hr, respectively, were: 1,080 and 133.1; 876 and 127.8; 818 and 129.6; 862 and 128.7.

The total units and the specific activities for the 12 enzymes assayed in the supernatants we give in Table 1, as are the percentage changes in specific activities between the six combinations of incubation times. Percentage change was apparently an unreliable indicator of a regulatory role for a given enzyme at a particular developmental stage. For example, isocitrate lyase increased in specific activity during each interval more than any other enzyme assayed, suggesting the operation of the glyoxalate bypass during those intervals; however, the preceding enzyme in the pathway, aconitase, decreased in specific activity in all but one interval, in which a slight increase occurred. Generally consistent trends were also not apparent in specific activities of other enzymes which could be pathway-related, such as fumarate and malate dehydrogenase; urease and the two glutamic dehydrogenases; and invertase, trehalase, and glucose-6-phosphate dehydrogenase.

Although it was known that enzymatic activities *in vitro* may have little relationship to enzymatic activities *in vivo*, it was nevertheless anticipated that consistent trends might be found owing to our use of synchronous cultures and short incubation times. Even under these conditions, however, enzyme profiles seemed to have little functional significance, as previously found by others (e.g., Hess and Brand 1965 *In Control of energy metabolism*, Chance et al., (Eds.), Academic Press, New York.

The following enzymes have been investigated previously during development of conidia: alcohol dehydrogenase and glucose-6-phosphate dehydrogenase (Weiss and Turian 1966 *J. Gen. Microbiol.* 44: 407), invertase and trehalase (Hanks and Sussman 1969 *Am. J. Botany* 56: 1152), isocitrate lyase (Turian et al. 1962 *Pathol. Microbiol.* 25: 737; Flavell

and Sanwal 1961 *Can. J. Microbiol.* 7: 319; Stine 1968 *J. Cell Biol.* 37:81), and NADH-oxidase (Stine 1968). We have used our method for obtaining synchronously-developing mycelial pads (1971 *Biochem. Genet.* 5:521) in examining the profiles of 12 enzymes to determine changes in their specific activities during development of conidiophore and conidia.

**Table 1.** Total units and specific activities of 12 enzymes, extracted after four incubation times, and % change in specific activities.

Enzyme	Total Units				Specific activity				% change in specific activity					
	Incubation time (hours)				Incubation time (hours)				Combinations of incubation times					
	0	1	4	7	0	1	4	7	0-1	0-4	0-7	1-4	1-7	4-7
<b>Aconitase</b>	4.85	3.11	3.32	2.77	36.4	24.4	25.6	21.5	-33	-30	-41	<b>+5</b>	-12	-16
<b>Acetaldehyde dehydr.</b>	26.6	22.4	17.7	13.0	200	176	<b>136</b>	<b>101</b>	-12	-32	-49	-22	-42	-26
<b>Fumarase</b>	1.30	1.20	1.20	<b>1.20</b>	9.7	9.4	9.3	9.3	-3	-4	-4	-1	-1	0
G-6-P dehydr.	58.9	49.2	49.2	55.0	442	<b>385</b>	380	427	-13	-14	-3	-1	<b>+11</b>	<b>+13</b>
<b>Invertase</b>	25.2	22.8	26.4	25.9	<b>189</b>	<b>179</b>	204	201	-5	<b>+8</b>	<b>+7</b>	<b>+14</b>	<b>+13</b>	-1
<b>Isocitrate lyase</b>	0.87	0.98	1.53	2.39	6.5	7.7	11.8	18.6	<b>+18</b>	<b>+82</b>	<b>+186</b>	<b>+53</b>	<b>+142</b>	<b>+58</b>
<b>Malate dehydr.</b>	0.51	0.47	0.65	0.58	3.8	3.7	5.0	4.5	-3	<b>+32</b>	<b>+18</b>	<b>+35</b>	<b>+22</b>	-10
NAD-GDH	3.62	3.47	2.68	2.75	27.2	27.2	20.7	21.4	0	-24	-21	-24	-21	<b>+3</b>
NADP-GDH	107	113	<b>113</b>	<b>116</b>	809	888	871	999	<b>+10</b>	+a	<b>+11</b>	-2	<b>+1</b>	<b>+3</b>
NADH-oxidase	2.03	<b>1.98</b>	1.91	1.23	15.3	15.5	14.8	9.6	<b>+1</b>	-3	-37	-5	-38	-35
<b>Trehalase</b>	4.11	3.69	3.27	3.13	30.9	28.9	25.2	24.3	-6	-18	-21	-13	-16	-4
<b>Urease</b>	22.4	18.7	21.3	20.6	168	147	164	160	-13	-2	-5	<b>+12</b>	<b>+9</b>	-2

• Expressed in millimicromoles of product/min/mg protein, except for **fumarase**, which is expressed as OD change/10 KC x 10<sup>6</sup>. All enzymes were assayed from the same extract for each incubation time.

• • • Department of Physiological Chemistry, University of Wisconsin, **Madison**, Wisconsin 53706 and Department of Microbiology, University of Georgia, Athens, **Georgia** 30601.